

JOURNÉE ANNUELLE DU CRPP 2025



«Diagnostic différentiel entre le PTI et les thrombopénies constitutionnelles » Point de vue de la recherche

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imagine
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Primary immune deficiency

Developmental, survival or functional defects of Immune cells

(T effector cells, regulatory T cells, B cells, neutrophils...)

caused by monogenic mutations in immune-related genes

(also called Inborn errors of Immunity, or IEI)

PIDs features:

Severe infections by
common pathogens

But also:

Cancers

Auto-immunity/inflammation/allergy



3 main functions

of the Immune System:

Clearance of pathogens

Tumor surveillance

Self-control

Monogenic mutations in immune related genes (>500)

can lead to

Primary Immune Deficiency and Dysregulation

(PIDD)

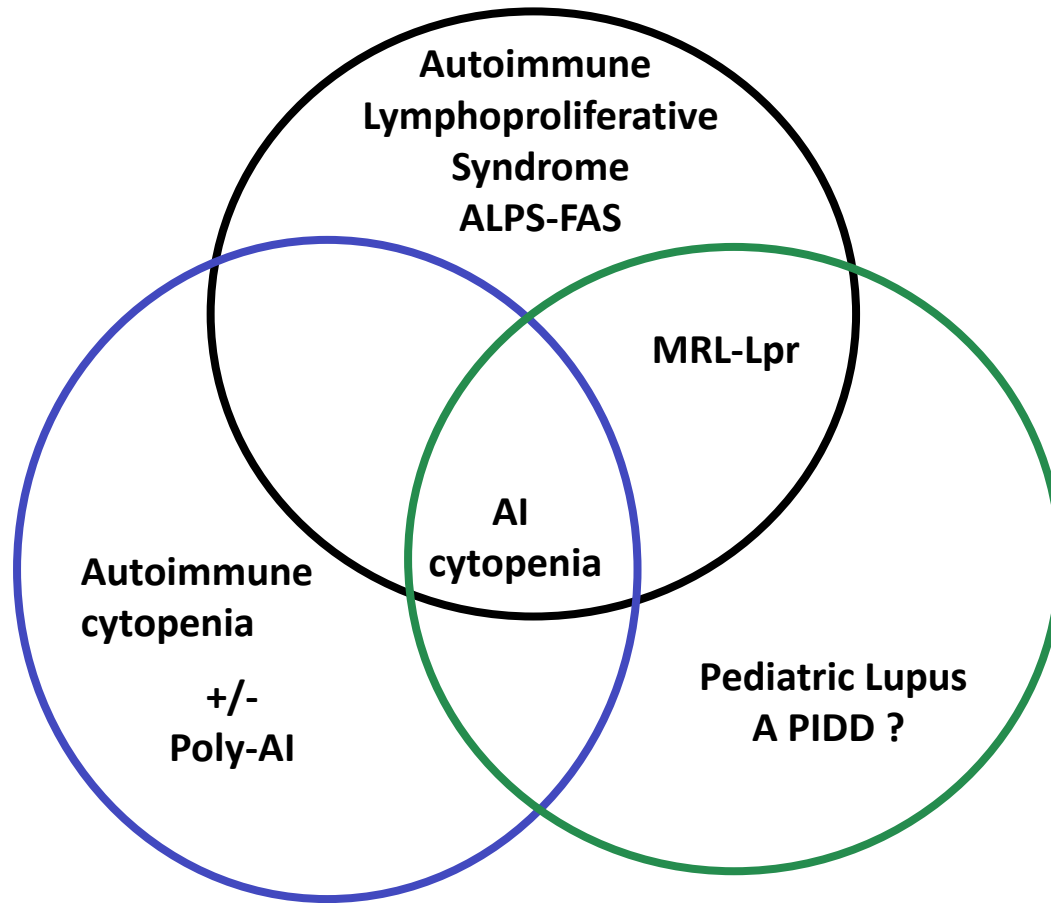
Relative risk (RR) of Autoimmunity in PIDD patients compared to the non-PIDD population

Register of >6000 PIDD cases (CEREDIH reference center: N.Mahlaoui A.Fischer)	Prevalence/ 1×10 ³ PIDD pts	Prevalence/ 1×10 ³ general population		RR
• Cytopenia	120	1		120
• Autoimmune hemolytic anemia (pediatric age)	25	.03	(France)	830
• Immune thrombocytopenia (adults)	60	1	(France)	60
• Inflammatory Bowel disease (adults)	78	1.8	(France)	43
• Inflammatory Bowel disease (pediatric age)	55	.7		80
• Skin disease	60	6	(France)	10
• Rheumatological disorders	50	8.6		6
• Rheumatoid arthritis (pediatric age)	8	.2	(France)	40

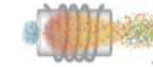
Autoimmune thrombocytopenia can be a presenting feature of PIDD

-> a rationale to search monogenic predisposition

30 years of research in Primary Immune Deficiency and Dysregulation (PIDDs)



Variants' impact validation



In-depth immunophenotyping (CyTOF)

Cellular models

Expression of variants in cell lines and primary cells



Cytokine production (Elisa, SIMOA)

iPSCs-> differentiation

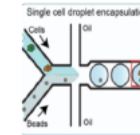
Functional and molecular assays



Autoantibodies

Animal models

Phenotype analysis treatments

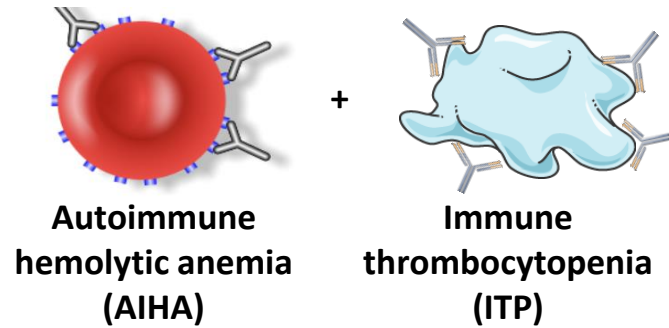


Single cell RNAseq, ATACseq

Implementation of multi-Omics analyses



Early onset autoimmune cytopenia: all monogenic diseases ?



Jérôme Hadjadj et al., Blood, 2019

M+ group 65%

Mainly heterozygous missense mutations

M- group 35%

Gene (no. of patients)	Mutation type and consequences
Pathogenic mutations, n = 32, 40%	
<i>TNFRSF6</i> (6)	Heterozygous/LOF
<i>CTLA4</i> (8)	Heterozygous/LOF
<i>STAT3</i> (6)	Heterozygous/GOF
<i>PIK3CD</i> (1)	Heterozygous/GOF
<i>CBL</i> (1)	Heterozygous/LOF
<i>ADAR1</i> (1)	Heterozygous/LOF
<i>LRBA</i> (4)	Homozygous/LOF
<i>RAG 1</i> (2)	Compound heterozygous/LOF
<i>TNFRSF6</i> somatic (1)	Heterozygous/LOF
<i>KRAS</i> somatic (2)	Heterozygous/GOF
Probably pathogenic mutations, n = 20, 25%	
Immune cell receptors	
<i>IFNAR1</i> (1)	Homozygous/likely LOF
<i>TNFR2</i> (1)	Heterozygous/likely GOF
<i>TGFB2</i> (1)	Heterozygous/likely LOF
Intracellular signaling	
<i>JAK1</i> (2)	Heterozygous/likely GOF
<i>JAK2</i> (1)	Heterozygous/likely GOF
<i>PLCG2</i> (1)	Heterozygous/likely GOF
<i>TRAF3</i> (1)	Heterozygous/likely GOF
<i>CARD11</i> (1)	Heterozygous/likely GOF
<i>ARHGEF4</i> (1)	Heterozygous/likely GOF
<i>PTPN11</i> (1)	Heterozygous/likely GOF
<i>PARP4</i> (1)	Compound heterozygous/likely LOF
Apoptosis regulation	
<i>RIPK2</i> (2)	Heterozygous/likely LOF
<i>APAF1</i> (1)	Heterozygous/likely GOF
Transcription factors	
<i>IKZF1</i> (2)	Heterozygous/likely GOF
<i>NFATC1</i> (2)	Heterozygous/likely GOF
<i>IKZF2</i> (1)	Heterozygous/likely LOF
No genetic abnormalities, n = 28, 35%	

Key pathways:

Immune Checkpoints
(CTLA-4/LRBA)

JAK/STAT
(JAK1/2gof, SOCS1/PTPN2lof)

Apoptosis
(FASLG/FAS/FADD)

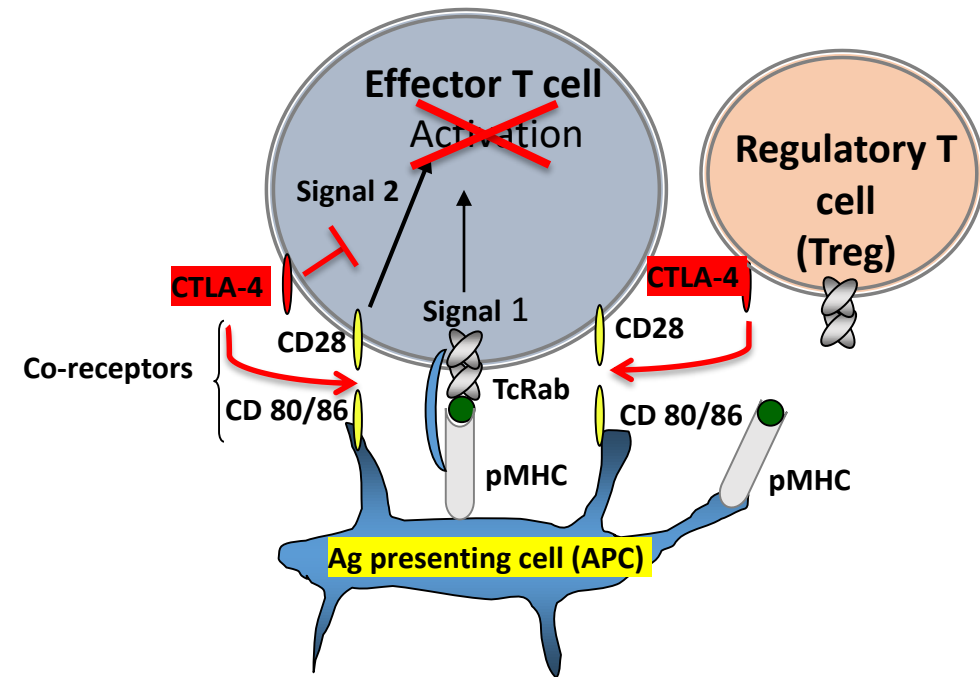
NFkB
(NFkB1/2, RELA, RELB, TNFAIP3)

MTOR
(PIK3CD, R1, ...)

Actinopathies
(ARPC1B, WAS, DOCK11, ACTN1...)

Autoimmune cytopenia associated with Immune checkpoint deficiency

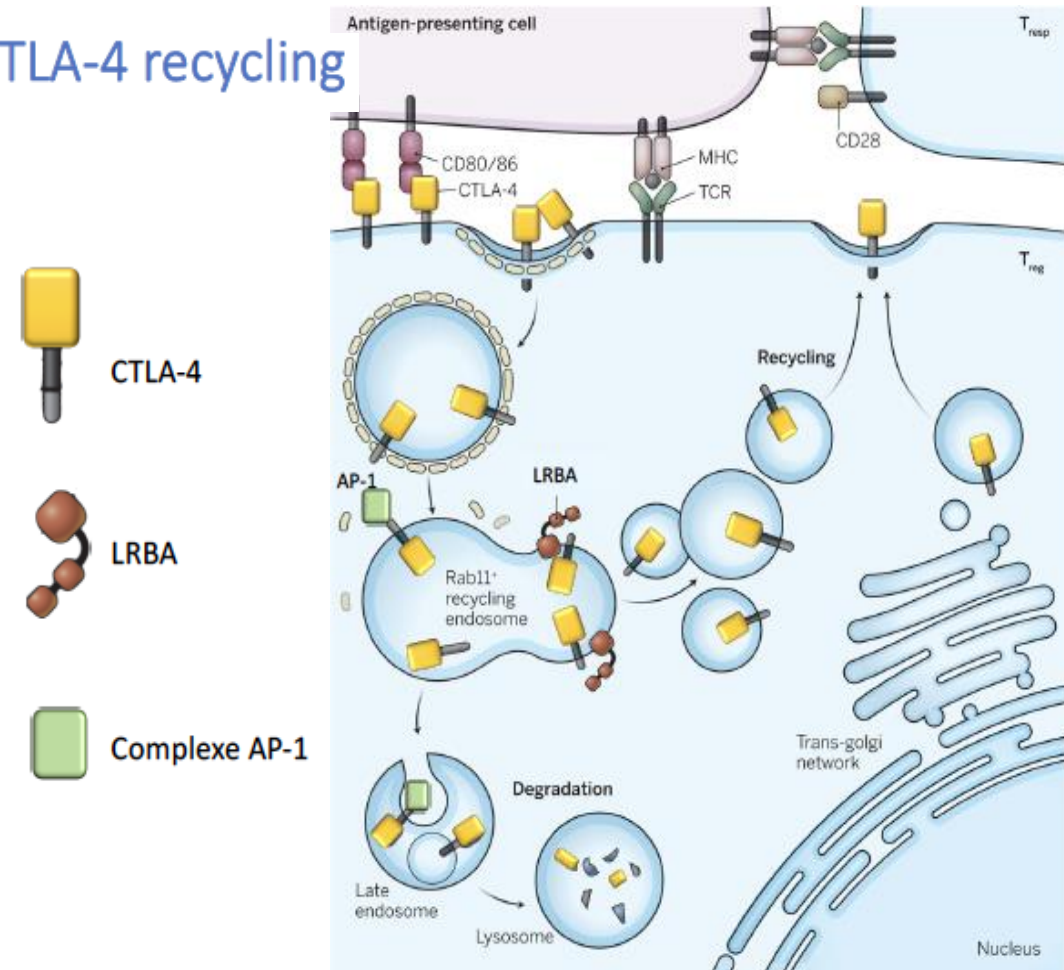
Role of CTLA4 and LRBA in effector and regulatory T cells



Inducible CTLA4 expression
in conventional T cells

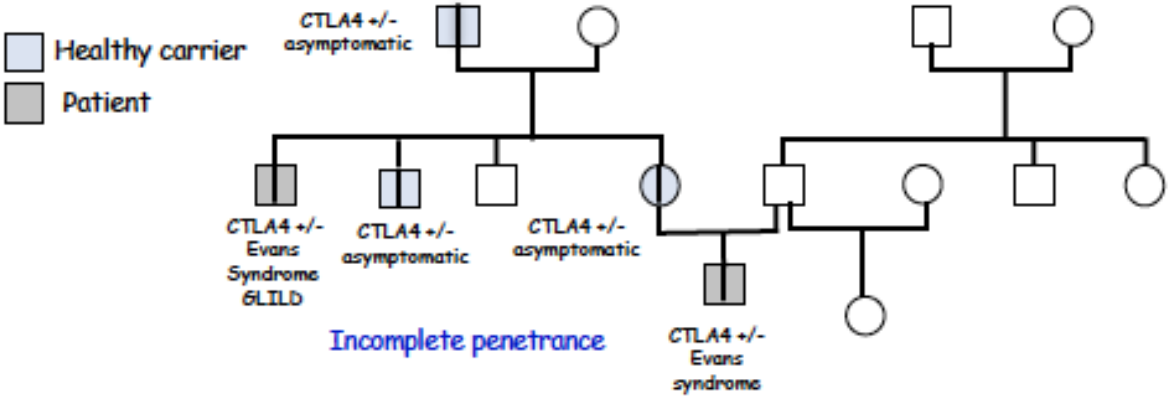
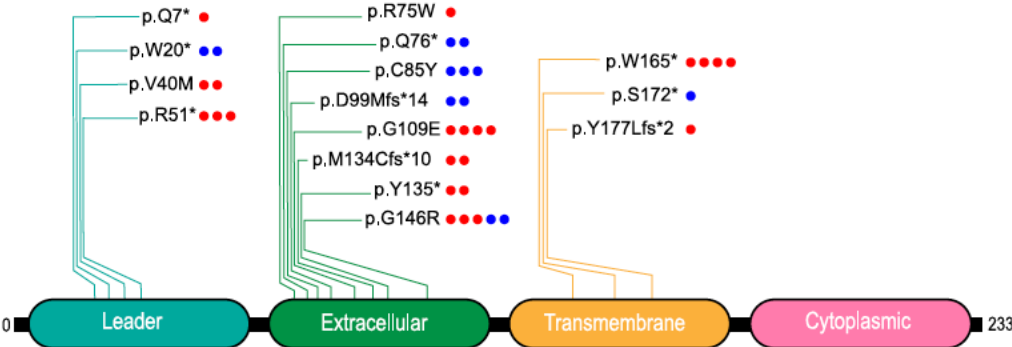
Constitutive CTLA4 expression
in Tregs

CTLA-4 recycling

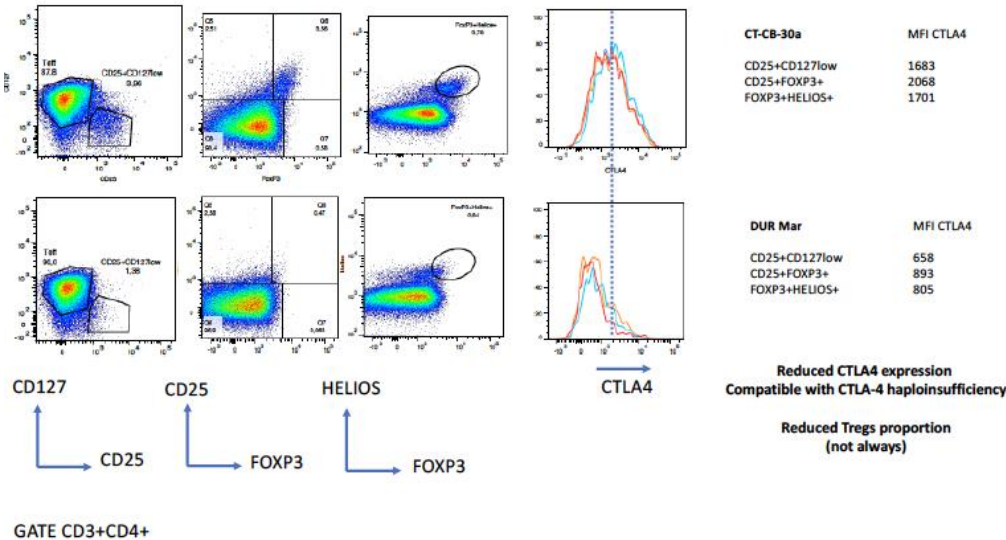


Autosomal dominant *CTLA-4* mutations with variable clinical penetrance

Exons 1-4 deletion

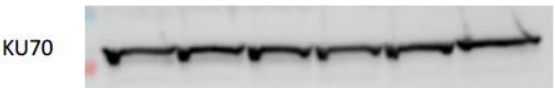
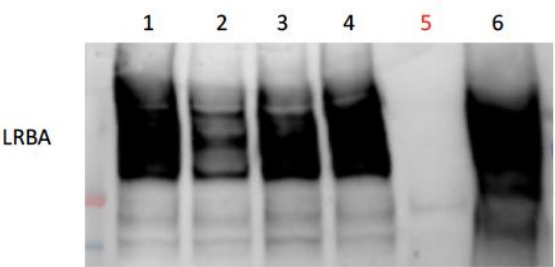
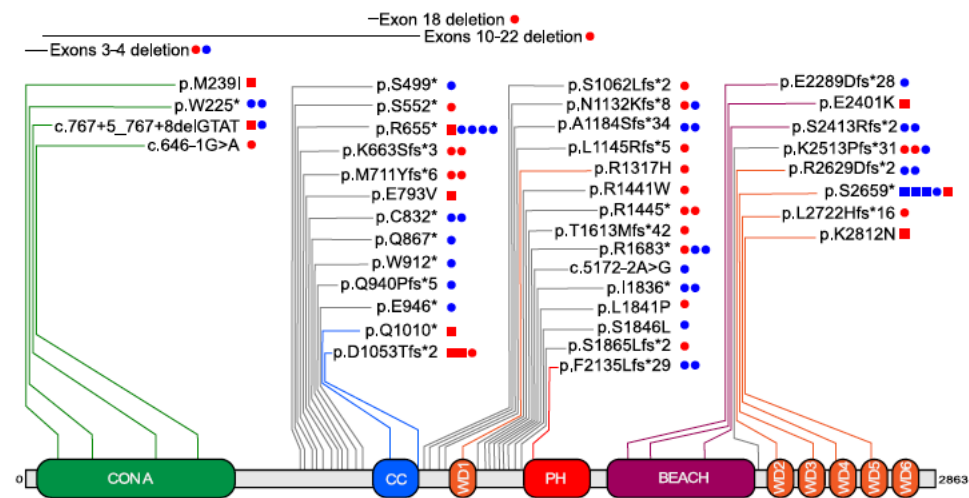


CTLA-4 mutations variable clinical penetrance

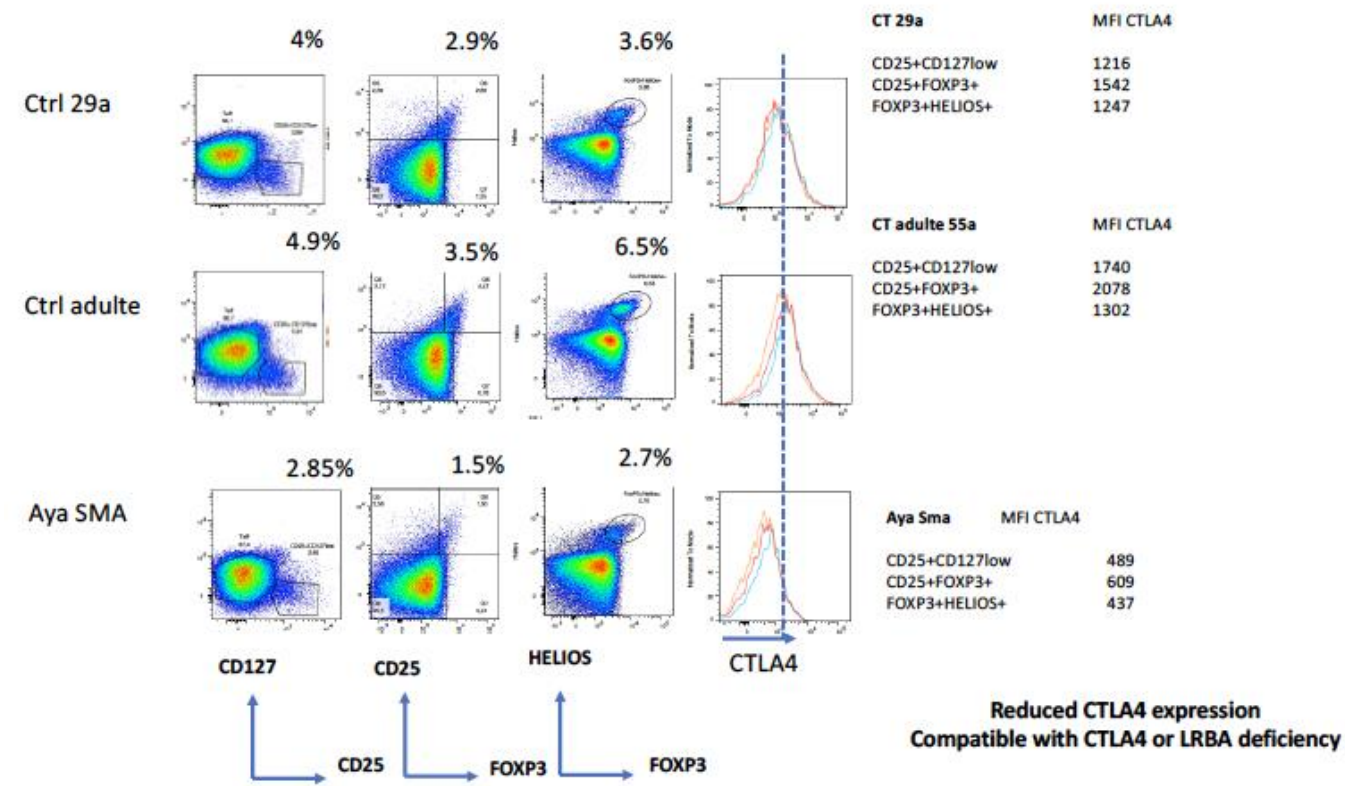


Membrane Staining
intracellular Staining

Autosomal recessive LRBA mutations: variable clinical manifestations



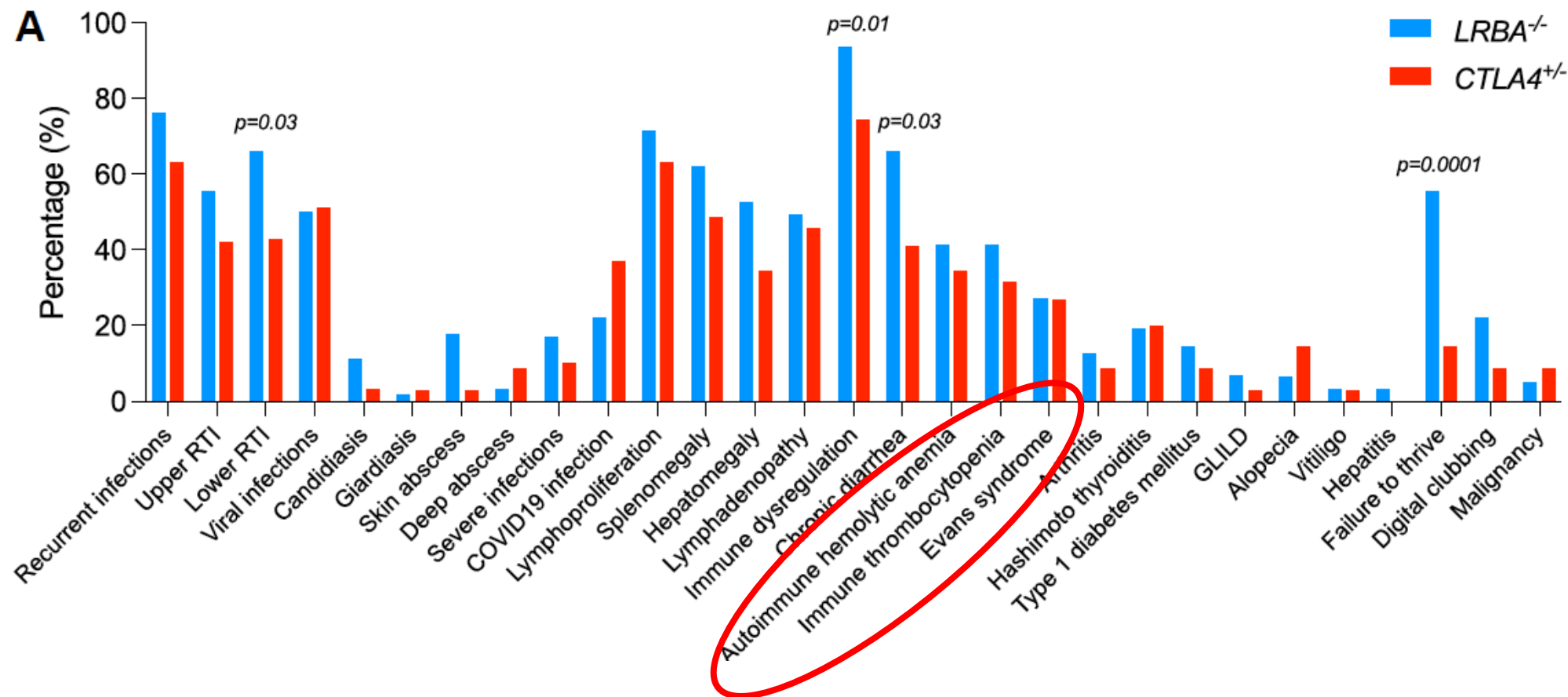
1= Ctrl Bs
2= suspicion
3= suspicion
4= suspicion
5= Sma Aya
6= Ctrl MJ



1-Reduced CTLA-4 expression on Tregs

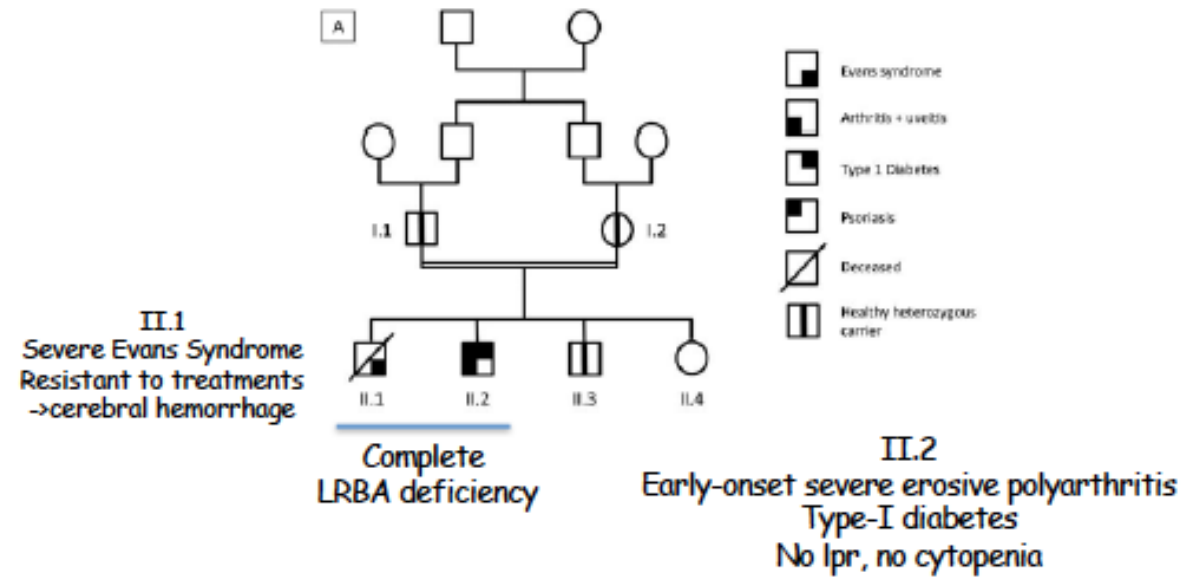
2-Absent (or profoundly reduced) protein expression on WB
(or intracellular staining)

Variable clinical expression in CTLA-4/LRBA deficiencies

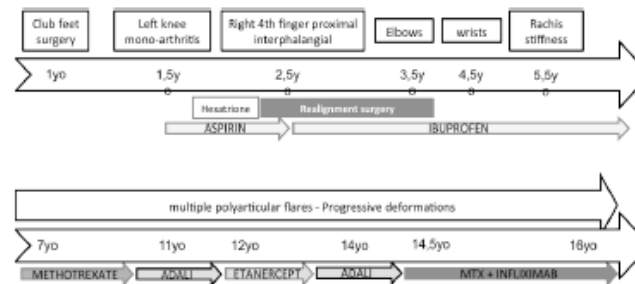
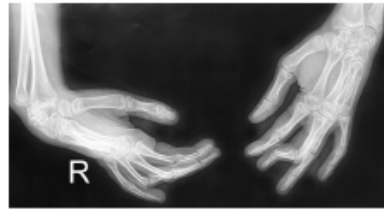


**Autoimmune cytopenia in 40-50% of patients with CTLA-4/LRBA deficiencies
and could be a presenting clinical feature**

Variable clinical expression autosomal recessive LRBA deficiencies

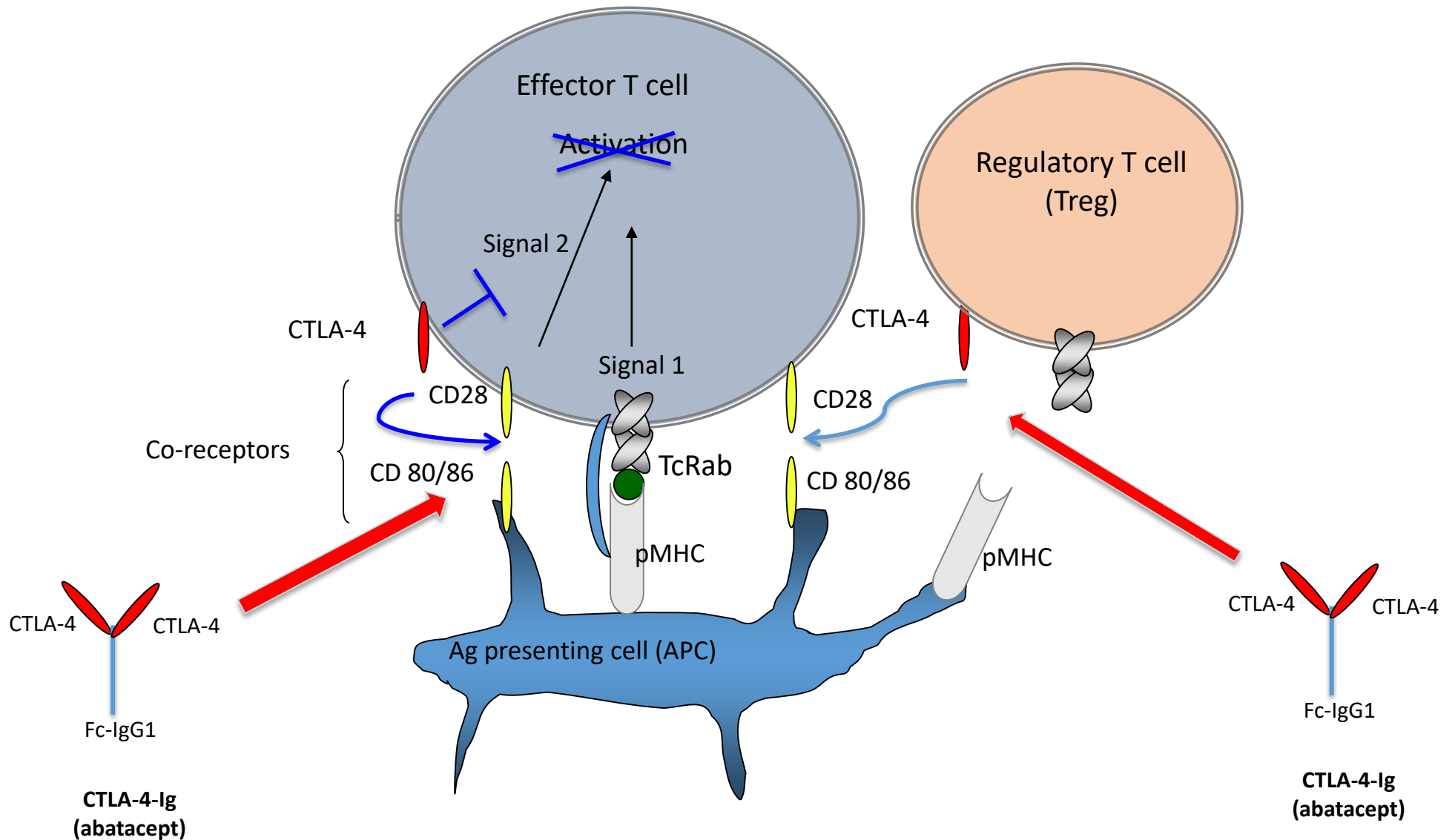


Patient II.2

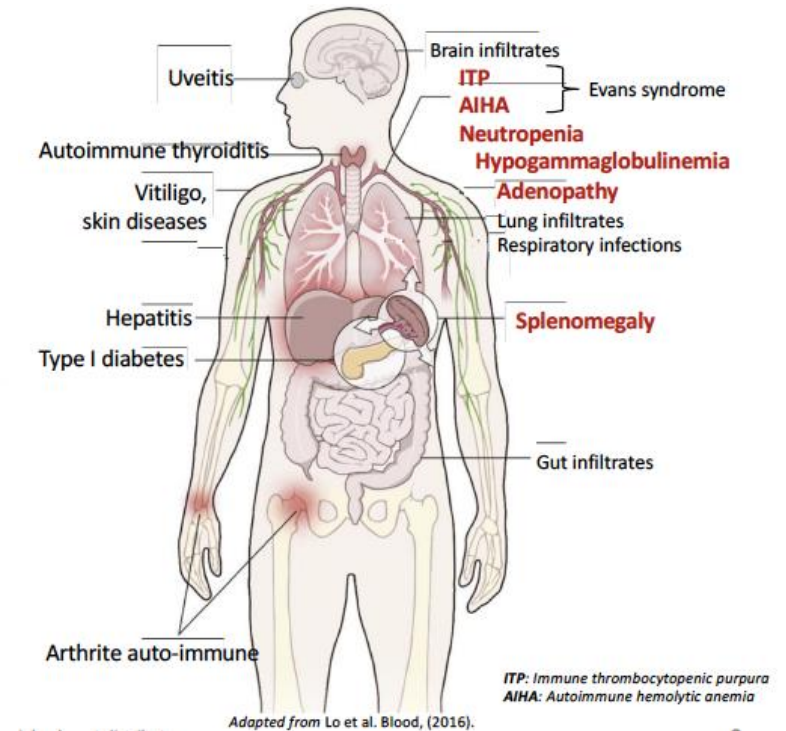
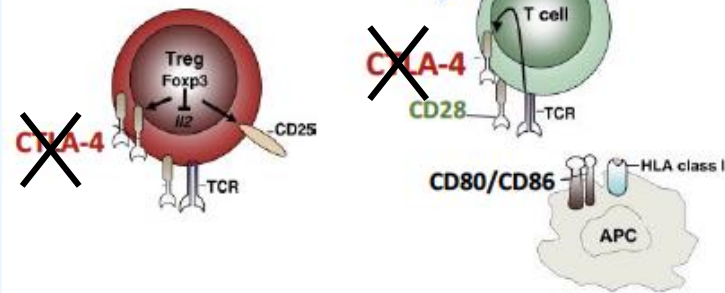
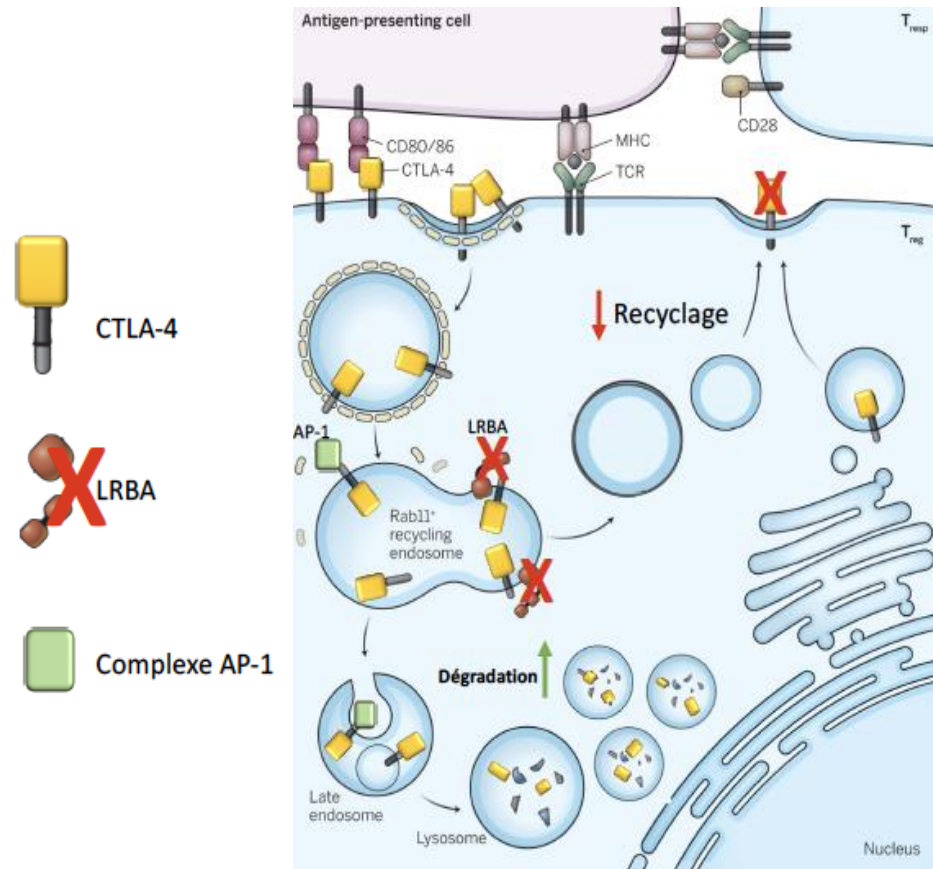


CTLA4 related pathophysiology
→ Targeted treatment

Role of CTLA4 in effector and regulatory T cells: towards targeted therapeutics



CTLA-4 or LRBA deficiencies are associated with low CTLA-4 expression in conventional T cells and Tregs



CTLA-4 haplo-insufficiency and LRBA deficiency are genetic phenocopies with low CTLA-4 expression on activated T cells and Tregs leading to various severe early-onset autoimmunity

Autoimmune LymphoProliferative Syndrome with mono-allelic *FAS* mutations: ALPS-FAS

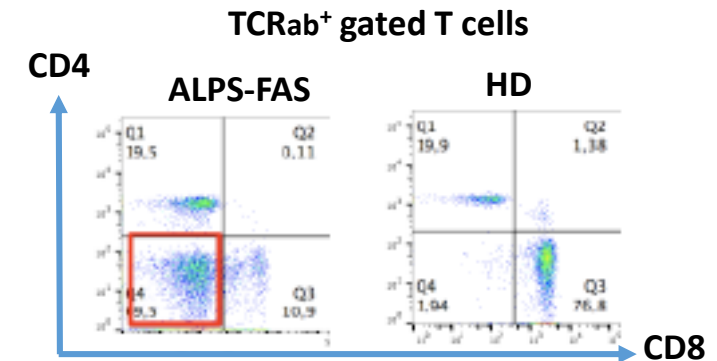
Chronic
Non-infectious
Non-malignant
Lymphoproliferation

Onset < 5 years (0-18y)
Splenomegaly (hepatomegaly)
Adenopathy



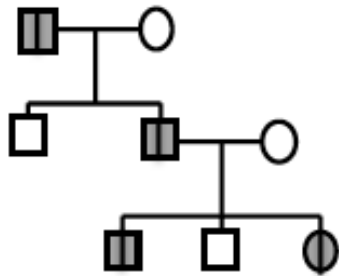
Immunological
disorders

«double negative» T cells (Tcrab⁺ CD4⁻ CD8⁻)
hyper Ig (G,A)
Elevated Vit B12, IL10, soluble FASL in sera
Autoimmune cytopenias (2/3 patients): ITP/AIHA
Rare organ-specific AI

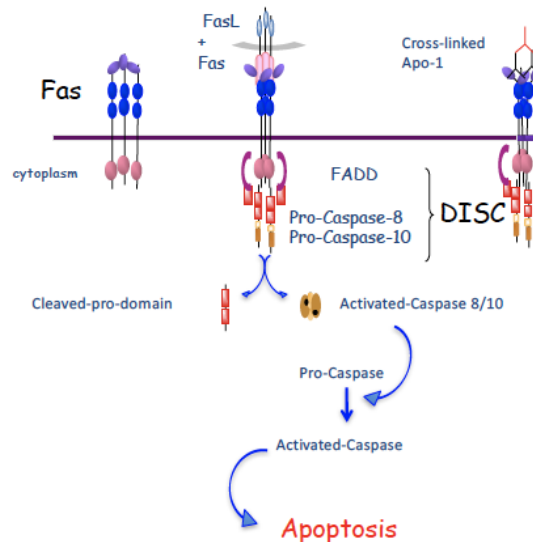


Familial cases

Dominant germline *FAS* mutations



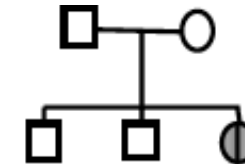
(Rieux-Laucat et al Science 1995)
(Fisher et al Cell 1995)



Impairment of *FAS*-induced apoptosis

Sporadic cases

Dominant somatic *FAS* mutations

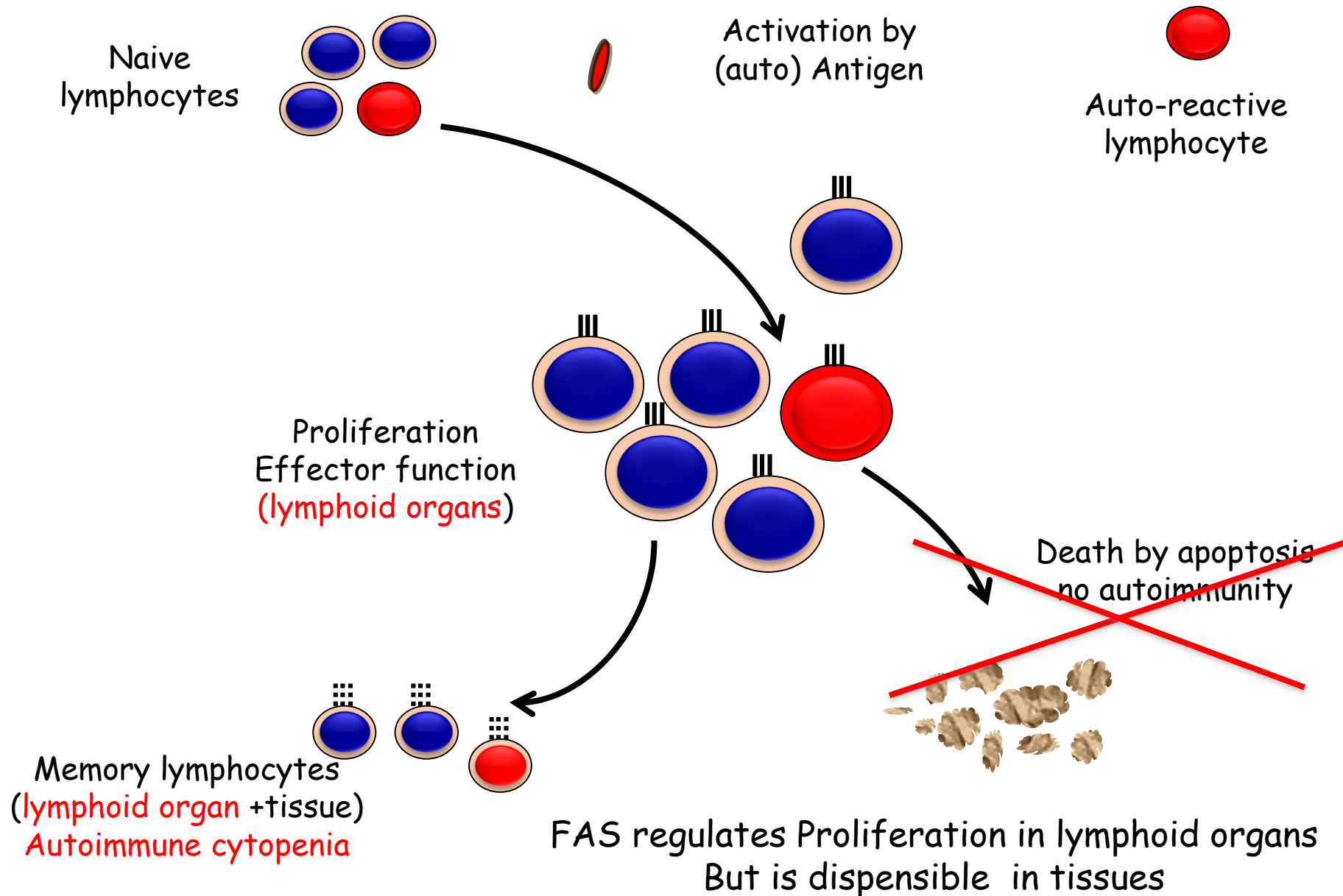


Somatic mutations appear
in hematopoietic precursors

Detected in DNT cells (80-100%)
in T and B cells , monocytes (1-20%)

(Holzelova et al NEJM 2004)

Programmed cell death deficiency: Lymphoproliferation + Auto-immunity



ALPS-FAS: differential diagnosis

- Découverte d'une thrombopénie à 106 000 puis 71 000 fin octobre 2004 à l'âge de 16 mois devant des lésions cutanées d'allure purpurique aux membres inférieurs. S'y associait une neutropénie autour de 500/mm³. Le myélogramme était riche en mégacaryocytes, la lignée granuleuse riche, avec un blocage partiel de maturation. Le bilan immunitaire était normal
- Myélogramme/caryotype médullaire/bilan d'hémostase/ échographie abdominale N,
- lymphopénie CD4 avec 850 lymphocytes/mm³ sans excès de lymphocytes T doubles négatifs.
- Conclusion: bilan en faveur d'une origine périphérique de la thrombopénie qui est à 70 000/mm³ avec 3 500 polynucléaires neutrophiles
- Absence de mutation du gène WAS en 2006 (eczéma et splénomégalie modérée)
- Consultation en octobre 2008: pas de syndrome hémorragique, 100 000 plaquettes, 4 000 polynucléaires neutrophiles.
- l'aspect des plaquettes sur le frottis sanguin n'a jamais montré à priori de particularité évocatrice d'un diagnostic précis, le volume plaquettaire moyen est normal, on peut retenir l'hypothèse d'un PTI chronique bien que l'évolution soit très atypique
- Dernière consultation en janvier 2015 : débord splénique de 4 cm, 58 000 plaquettes, 2 000 PN, 1600 lymphocytes, 560 CD4, CD4- CD8- 3 %, PCR EBV négative, FAS ligand 0,6 ng/mL, IL10 0, vitamine B12 891 pg/mL pour une normale entre 180 et 820, test de Coombs négatif : IgG 12,5 g/L, IgA 1,49 g/L, IgM 4,2 g/L, sérologie EBV positive en IgG VCA, EBNA et IgM VCA

-> NGS (research)

-> Homozygous mutation in *NBEAL2*

Gray platelet syndrome can mimic autoimmune lymphoproliferative syndrome

[Anne Rensing-Ehl](#), [Ulrich Pannicke](#), [Stefanie-Yvonne Zimmermann](#), [Myriam Ricarda Lorenz](#), [Benedicte Neven](#), [Ilka Fuchs](#), [Ulrich Salzer](#), [Carsten Speckmann](#), [Anne Strauss](#), [Eberhard Maaß](#), [Benedicte Collet](#), [Anselm Enders](#), [Remi Favier](#), [Marie Christine Alessi](#), [Frederic Rieux-Laucat](#), [Barbara Zieger](#), [Klaus Schwarz](#), [Stephan Ehl](#)

Blood. 2015 Oct 15; 126(16): 1967–1969.

IgG g/L* (normal age range, y)	23.41 (5.04-14.74)	10.46 (4.53-9.16)	16.51 (5.04-14.74)	10.6 (6.98-15.84)	13.74 (6.98-15.84)	18.2 (5.04-14.74)
DNT %† (<2.5%)	2.8 (43/μL)	1.7 (24/μL)	2.4 (28/μL)	1.5 (14/μL)	2.3 (35/μL)	5 (57/μL)
Vitamin B ₁₂ ,*,‡ mg/dL	3 856	>4 000	>4 000	3 758	1 843	>2 000
sFASL,‡ pg/mL (<200)	759	740	797	549	465	630
IL-10, pg/mL (<20)	6.5	<1	4.7	<1	<1	<1

ITP, Splenomegaly, Hyper IgG, Increased soluble FAS-ligand

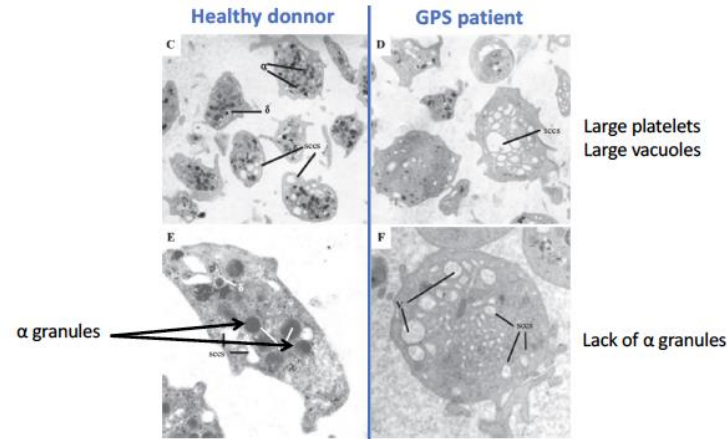
-> pathophysiology ??

Grey Platelet syndrome: a bleeding disorder with autoimmune features

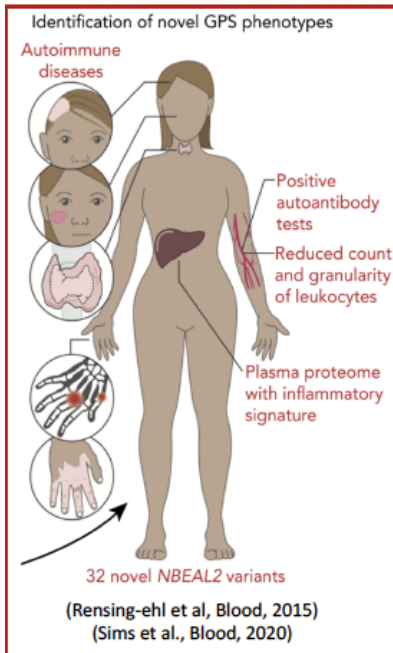
What are the mechanisms involved in autoimmunity?

Gray platelet syndrome (GPS)

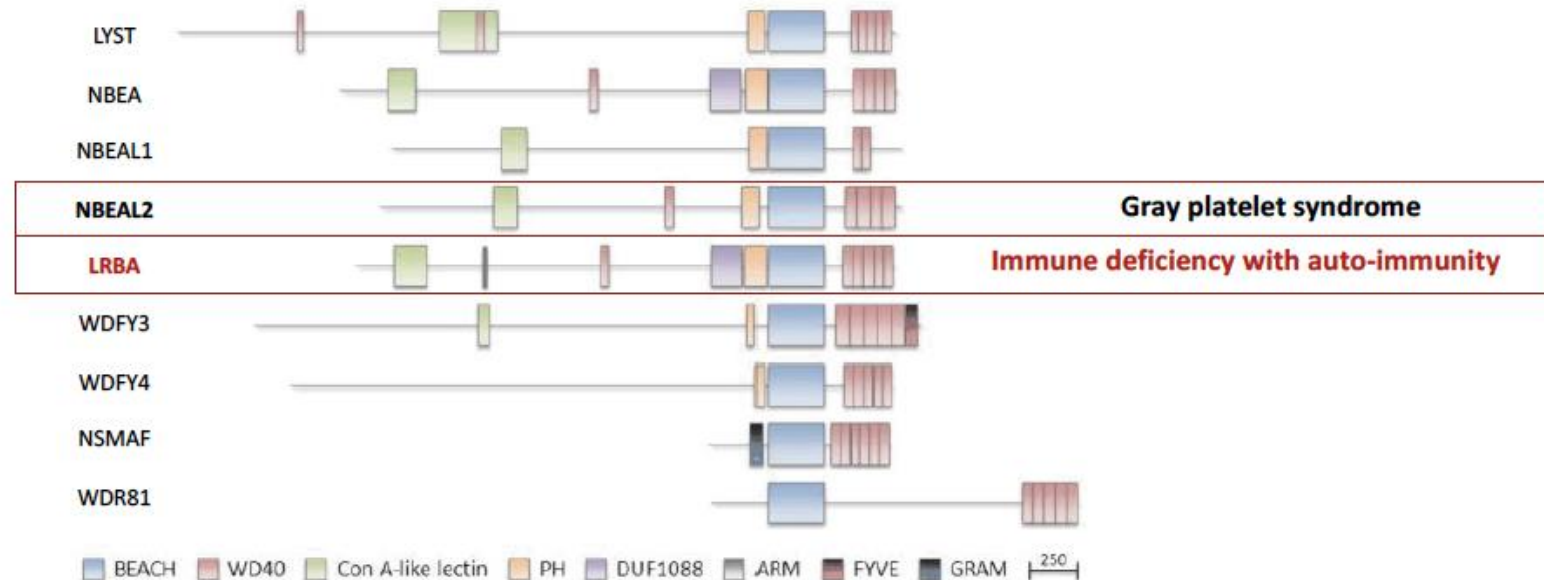
Congenital hemorrhagic syndrome caused by **recessive NBEAL2 mutations**
Abnormal gray platelets and macrothrombopenia
Often splenomegaly, sometimes evolving in progressive myelofibrosis



Adapted from Rosa, J. P. (2014). Sang Thrombose Vaisseaux



BEACH protein family



Gray platelet syndrome

Immune deficiency with auto-immunity

LRBA is involved in CTLA-4 trafficking through its BEACH domain

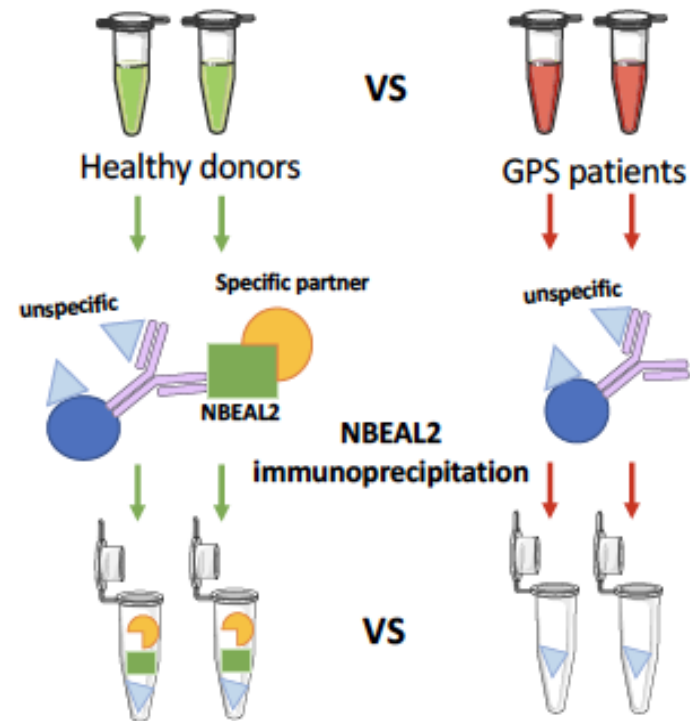
LRBA and NBEAL2 are structurally similar and involved in vesicular trafficking

Adapté de Cullinane et al. Traffic (2013)

Hypothesis:

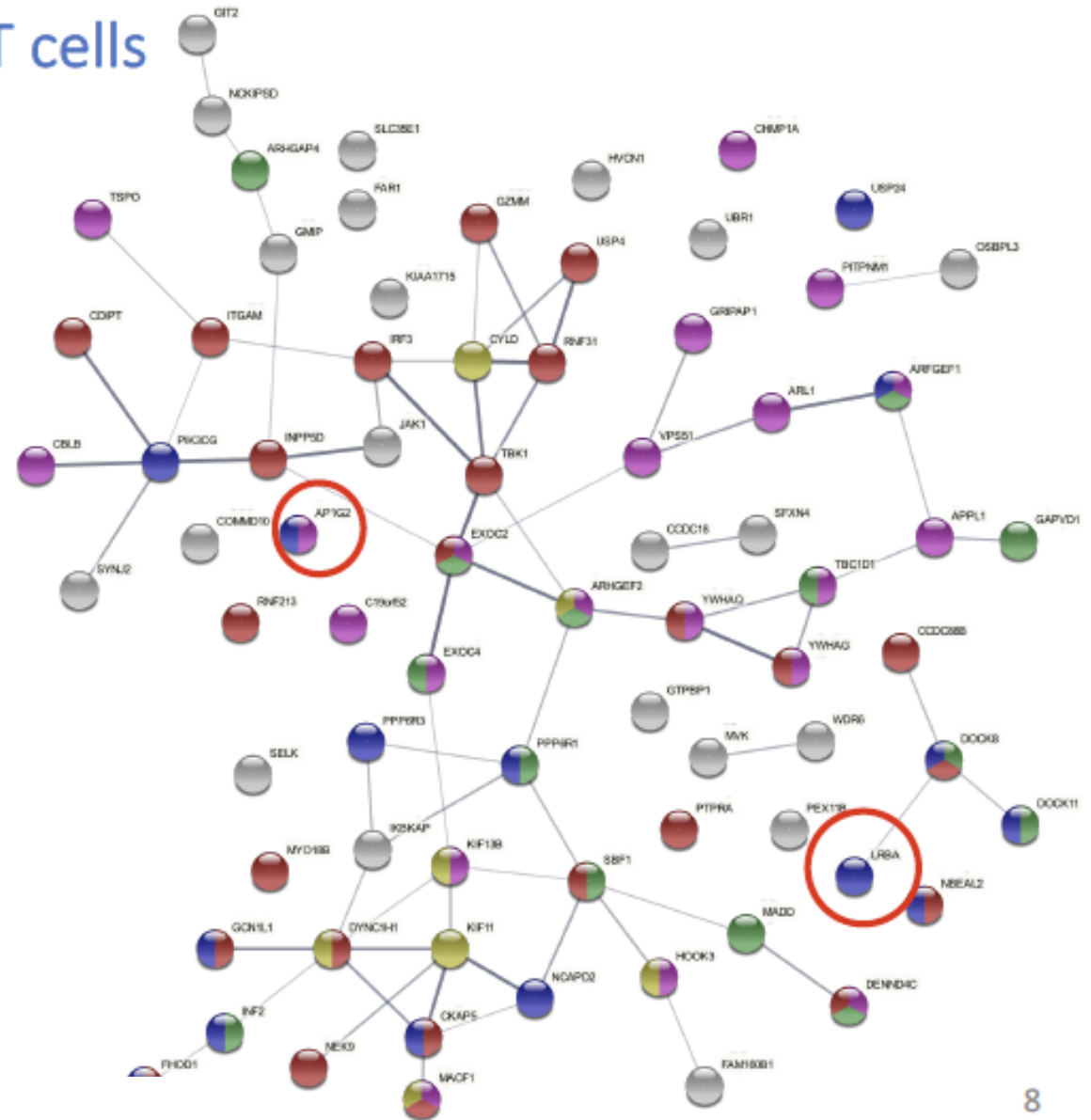
NBEAL2 is involved in the vesicular trafficking of immune receptors
(Laure Delage, Thèse CIFRE SANOFI)

NBEAL2 interactome in activated T cells

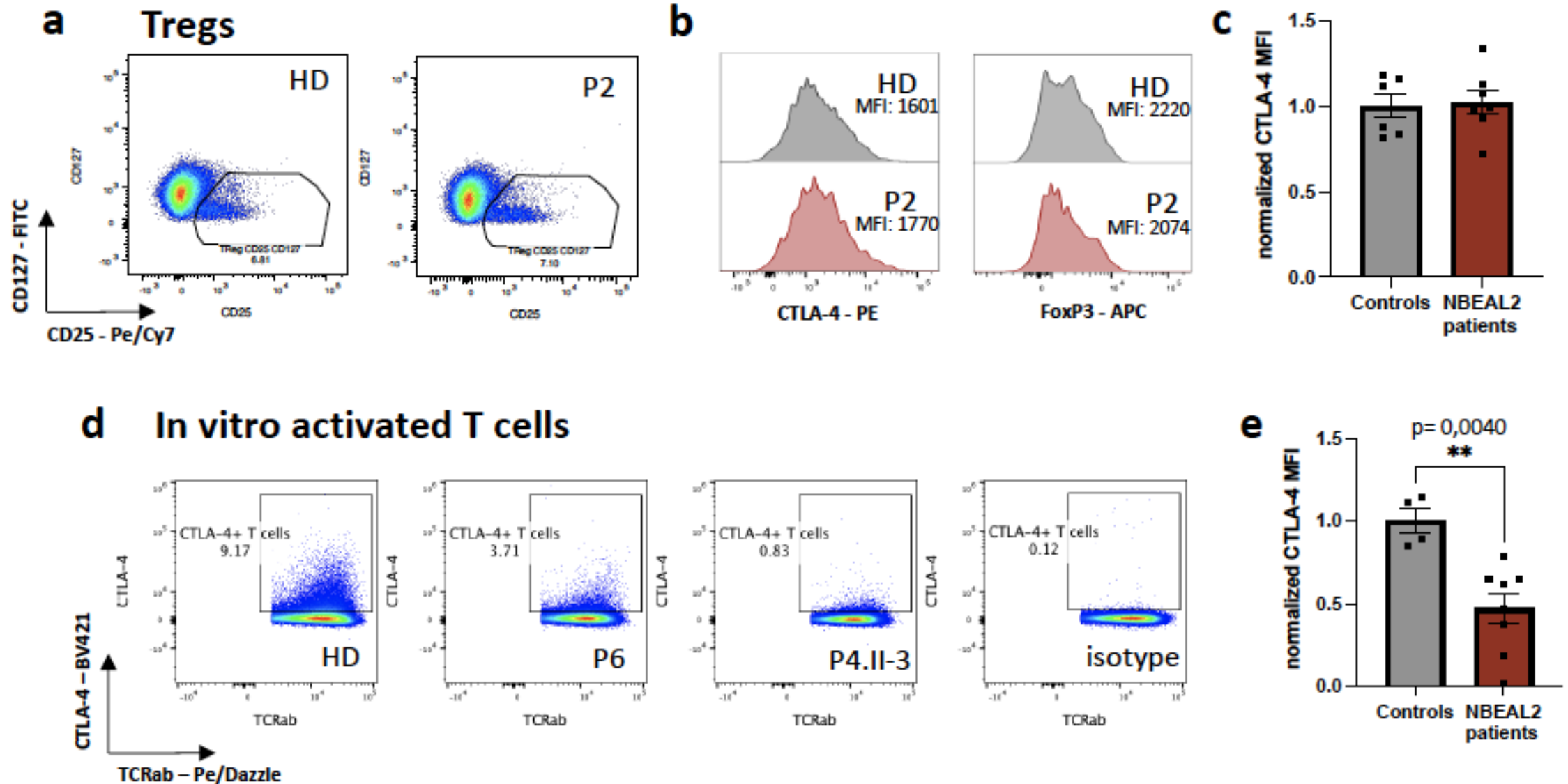


- | | | |
|---|---|---|
| ● Immune system
(25/76 proteins) | ● GTPase binding
(14/76 proteins) | ● ARM domain containing
(15/76 proteins) |
| ● Transport proteins
(21/76 proteins) | ● Microtubules
(7/76 proteins) | |

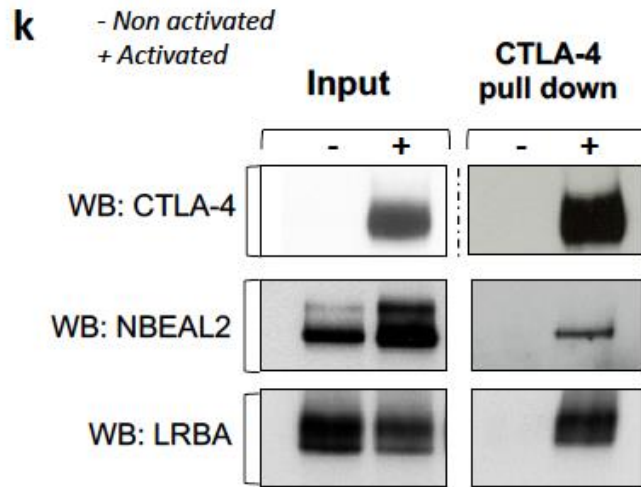
Proteins involved in CTLA-4 trafficking (LRBA, AP1G2)



NBEAL2 deficiency is associated with normal CTLA-4 expression in Tregs But low CTLA-4 expression in Activated conventional T cells

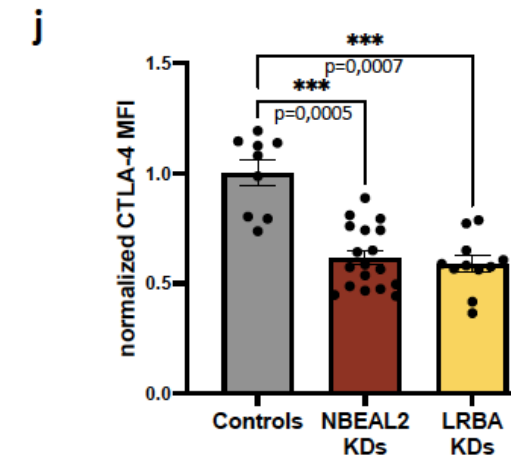
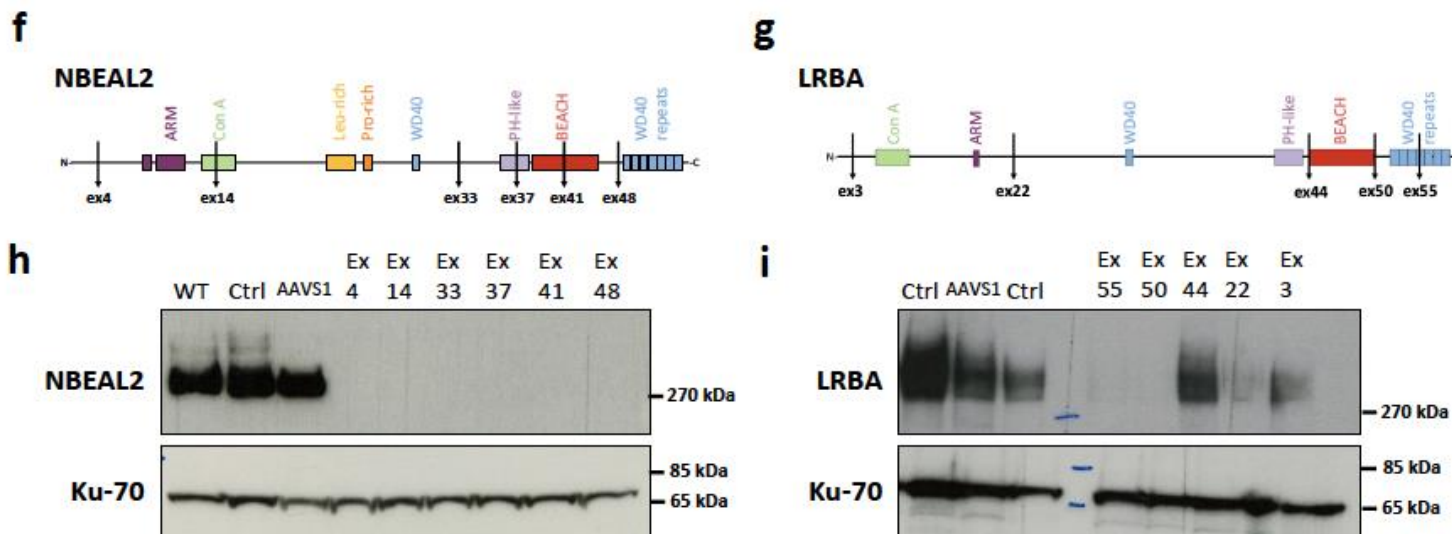


NBEAL2 and LRBA interact with CTLA-4 in primary T cells from healthy controls

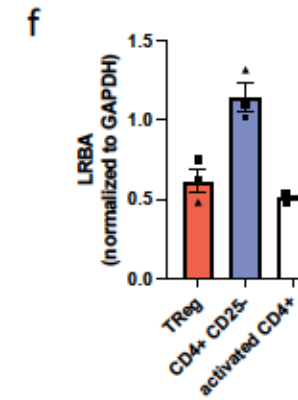
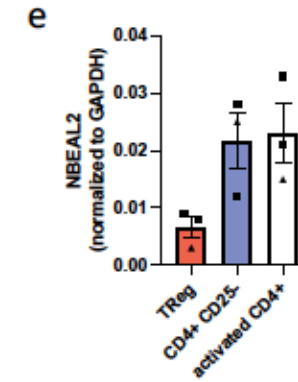
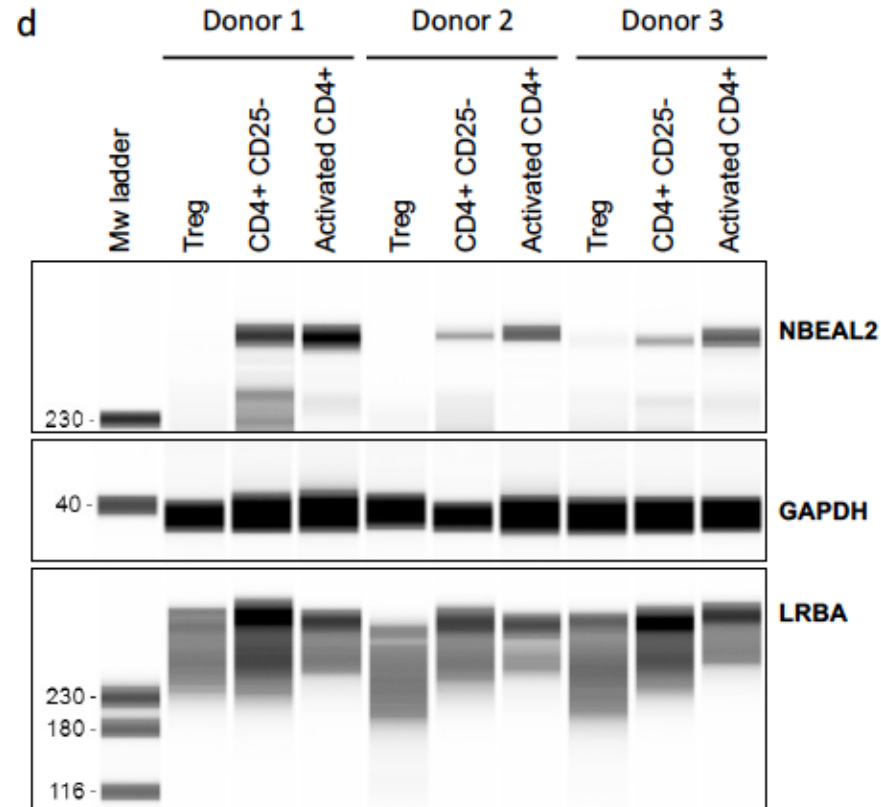


Pull-down of CTLA-4 reveals interaction with LRBA and NBEAL2 in primary activated T cells from healthy controls

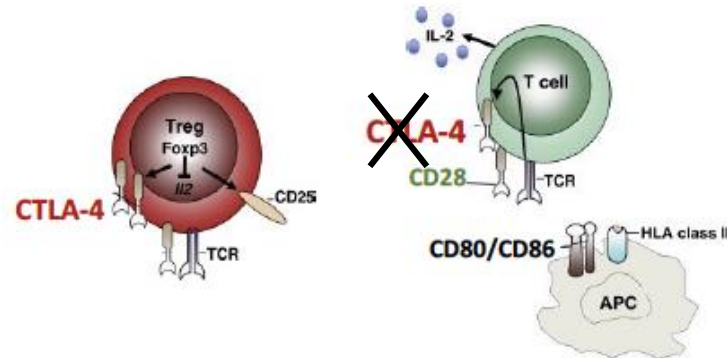
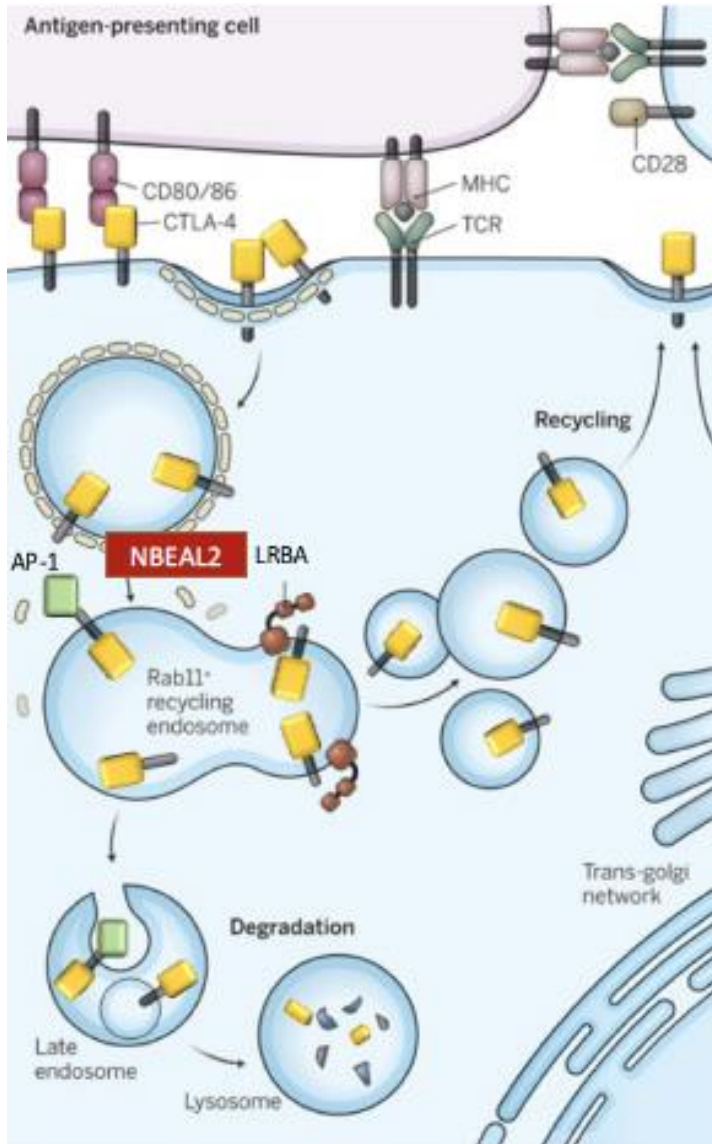
NBEAL2 or LRBA Knock-down by Crispr/Cas9 in primary T cells from healthy controls



NBEAL2 is expressed in conventional T cells but not in Tregs from healthy controls



NBEAL2 regulates the CTLA-4 expression in conventional activated T cells



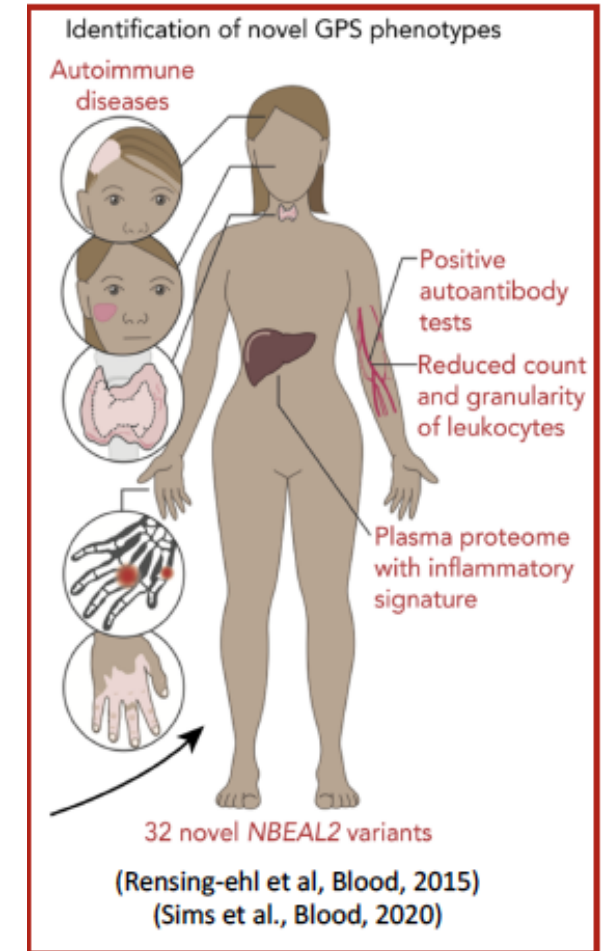
NBEAL2 :

interacts with LRBA

**Regulates CTLA-4 expression
on activated conventional T cells
but not on Tregs**

**Low CTLA-4 expression on activated T cells
Is associated with autoimmunity**

**-> Abatacept as a potential therapy
To treat autoimmunity in GPS?**



Constitutional Platelet deficiencies: A possible genetic cause of autoimmune cytopenia? Platelets target and culprit in ITP ?

mutations in **WAS**, **NBEAL2**, and genes associated with constitutional thrombocytopenia
in patients with Autoimmune cytopenia

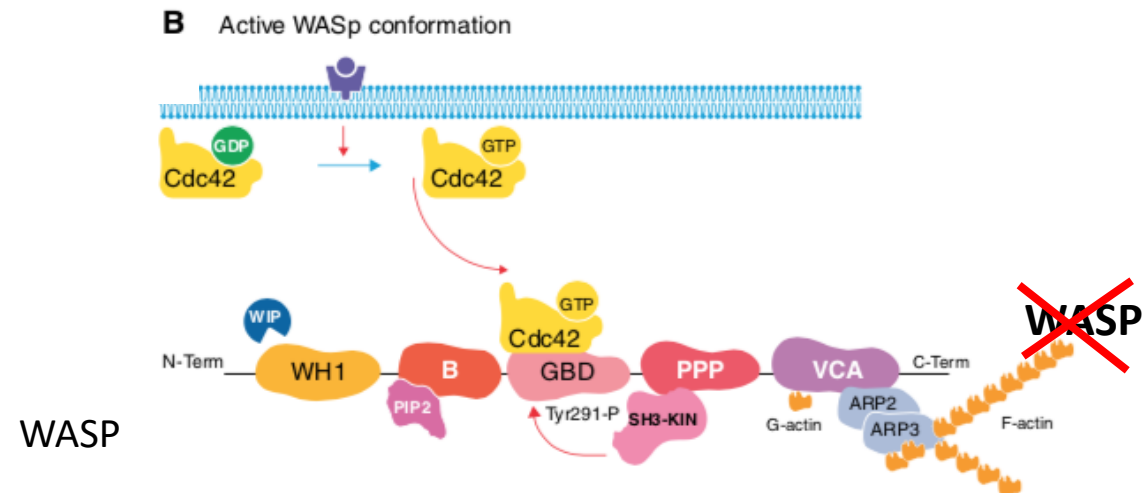
WAS hemizygote mutations:

-Aldrich syndrome:

X-linked CID, thrombocytopenia with microplatelets,
Autoimmunity

-XL thrombocytopenia

WASp KO in mice recapitulates the human disease



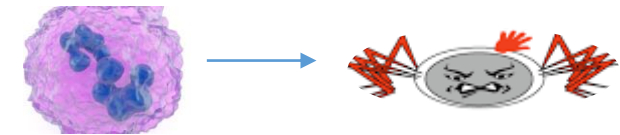
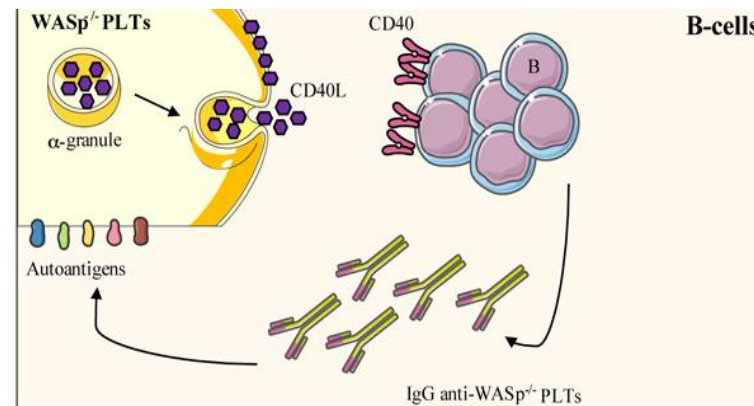
Mice WAS^{KO} in MK

Conditional WASp KO in megakaryocytes:

-> microplatelets

AND

Autoantibodies against platelets



**Abnormal platelets production
Can contribute to immune tolerance
breakdown**

Constitutional Platelet deficiencies: A possible genetic cause of autoimmune cytopenia?

Platelets target and culprit in ITP ?

mutations in WAS, NBEAL2, and genes associated with constitutional thrombocytopenia in patients with autoimmune cytopenia

Patients with AI cytopenia
and mutations in genes of constitutional thrombocytopenia

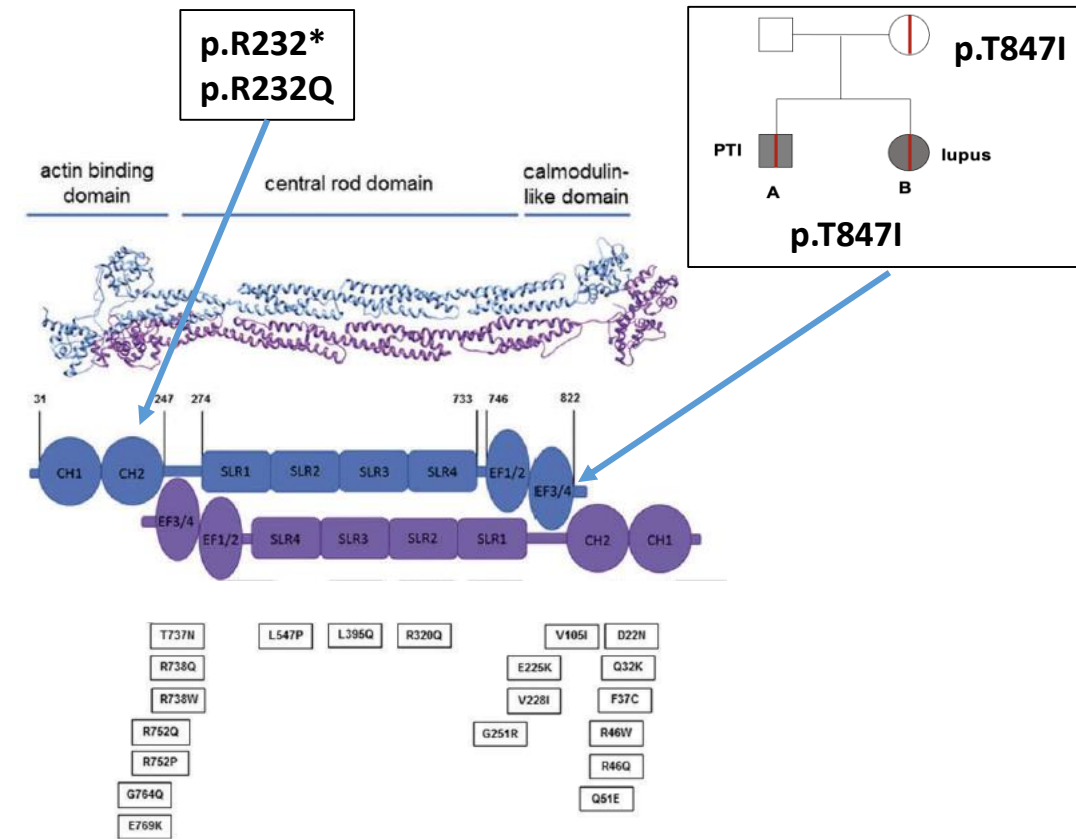
Platelet
function

	Gene	protéine	CADD	MAF
P1	TUBB1	p.R297H	28	Nd
P2	TUBB1	p.E45X	NA	Nd
P3	ACTN1	p.N462H	28	Nd
P4	TPM4	p.E75Q	24	Nd
P5	TRPM7	p.A1254V	23	4.80x10 ⁻⁴
P6	MYH9	p.D1424N	28	Nd
P7	MYH9	p.S1294N	23	Nd
P8	MYH9	p.D1627N	24	5.66x10 ⁻⁵
P9	MYH9	p.D1119V	27	Nd
P10	ANKRD26	p.R858X	38	Nd
P11	ETV6	p.R369W	34	Nd
P12	RUNX1	p.M267I	16	3.93 x10 ⁻⁵
P13	JAK2	p.F240V	23	Nd
P14	PLCG2	p.V431	22	2,14 x10 ⁻⁵
P15	PLCG2	p.D520G	21	3,21x10 ⁻⁵

Immune
and
Platelet
function

Validation ongoing (F.Zeco)

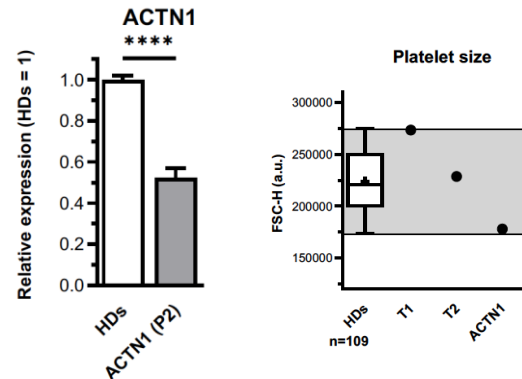
ACTN1 variants



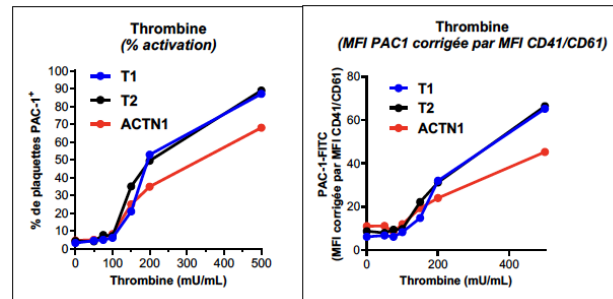
Mutations in constitutional thrombocytopenia

ACTN1 variants: A possible genetic cause of autoimmune cytopenia?

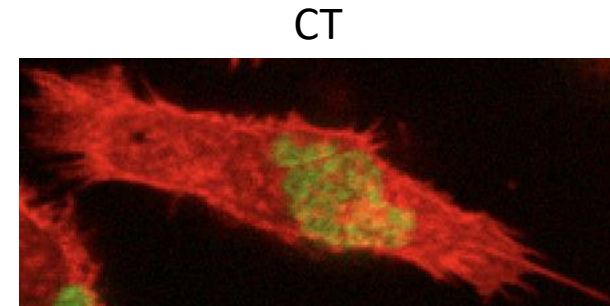
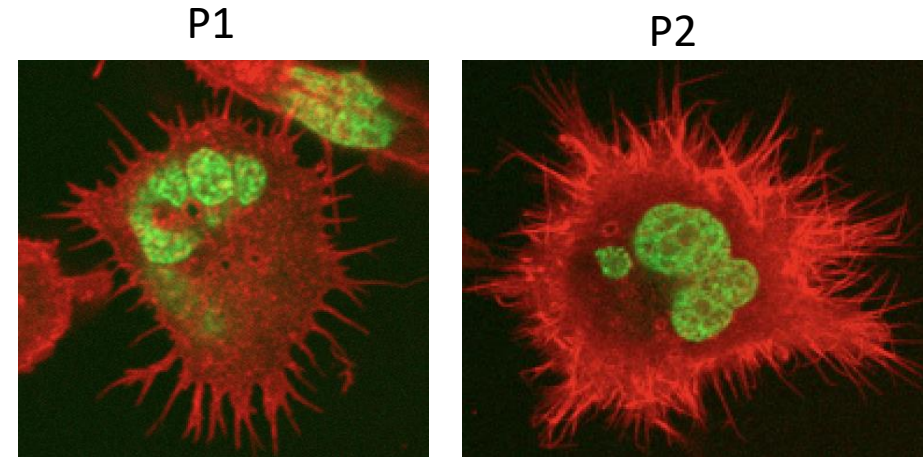
F.Zeco (ongoing PhD study), Coll: F. Adam and A Kauscot (UMRS-1176, KB)



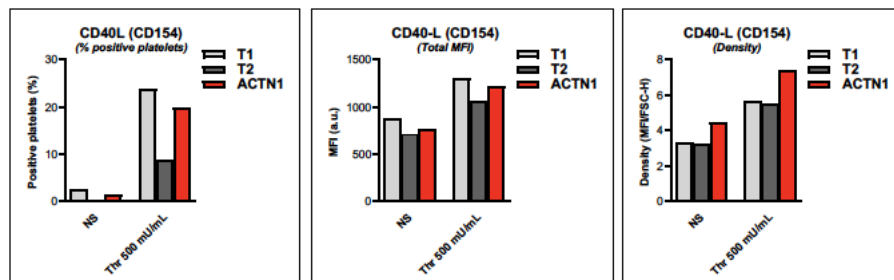
Reduced protein expression and platelets' size



Reduced platelet activation



Abnormal morphology in B-EBV cell lines

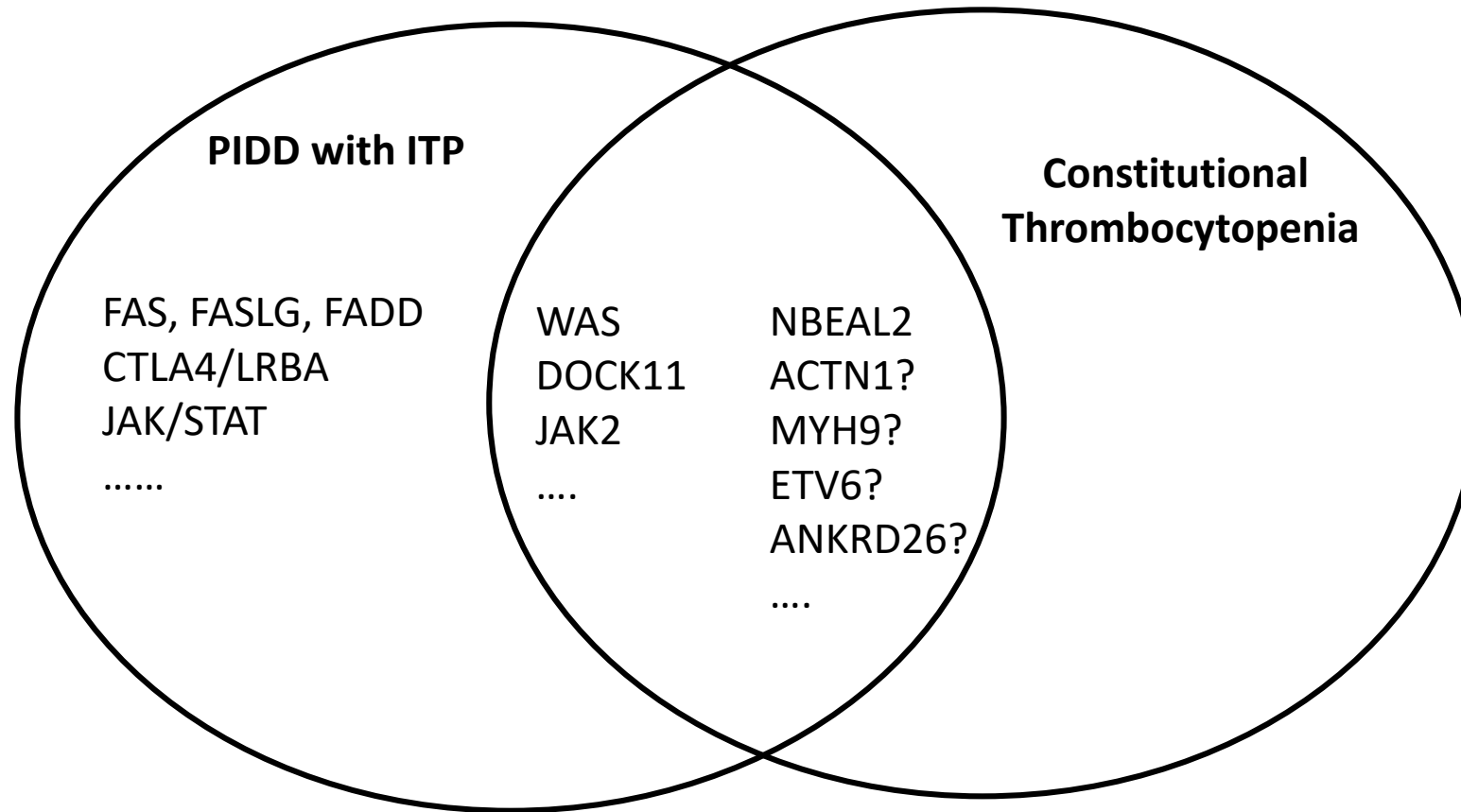


Increased CD40LG density

ACTN1 variants:
Preliminary results on platelets to be confirmed

Role of ACTN1 variants in lymphocytes ?
-> Tregs function (Trogocytosis), T/B cell functions

Constitutional Platelet deficiencies and ITP: Conclusions



Mutations in genes with dual roles in tolerance and platelet biogenesis (WAS, NBEAL2, DOCK11, JAK2...)

Abnormal platelets production/function can contribute to immune tolerance breackdown

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